

Evidence for a signal peptide at the amino-terminal end of human mitochondrial aldehyde dehydrogenase

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A full-length cDNA clone coding for human mitochondrial aldehyde dehydrogenase (ALDH I) was isolated from a human fetal muscle cDNA library. Sequence analysis revealed structural similarities between the amino-terminal end of ALDH I and other known targeting sequences responsible for protein uptake into the mitochondria.

Leader peptide; cDNA sequence; Aldehyde dehydrogenase I; (Human)

1. INTRODUCTION

Human liver aldehyde dehydrogenase (ALDH, EC 1.2.1.3) consists of at least four isozymes. The multiple forms of ALDH show a considerable heterogeneity in their tissue and organ distribution as well as subcellular localization [1,2]. ALDH I is also referred to in the literature as E₂ or ALDH2 isozyme [2]. Localization of this isoenzyme in the mitochondria as well as its assignment to chromosome 12 [3,4], suggests the existence of a leader peptide directing the protein uptake into the mitochondria. Most nuclear coded mitochondrial proteins are translated on cytoplasmic polysomes at larger precursors containing leader peptides of 20–60 residues in length at the amino-terminal end of the protein [5]. Usually these peptides are characterized by the absence of acidic residues and an average to above average content of basic residues periodically separated by uncharged residues [6,7]. Beside these general structural similarities, no significant sequence homologies were found between mitochondrial presequences.

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In a recent study, it has been shown that even a duplication of a part of the amino-terminal end ('cryptic mitochondrial targeting signal') of a cytosolic protein is sufficient to direct its incorporation into the mitochondria [8].

Previously, a partial cDNA clone pEXAL21 coding for the carboxy-terminal end of ALDH I was isolated and characterized from a human liver cDNA library [9,10]. Until now, no sequence data were available for the amino-terminal end of the ALDH I protein. The cDNA clone pEXAL21 was chosen to screen a human fetal muscle cDNA library in order to isolate a full-length cDNA clone and especially to obtain information on the extreme amino-terminal end.

2. MATERIALS AND METHODS

Human fetal muscle RNA was isolated from abortion material in the 22nd week of gestation. The cDNA library was established in the expression vector λ gt11. RNA isolation and cDNA synthesis were performed as in [11]. 2×10^6 pfu of the human fetal muscle cDNA library were screened using a nick-translated cDNA fragment derived from the ALDH I clone pEXAL21. DNA was isolated from positive λ clones by a small-scale

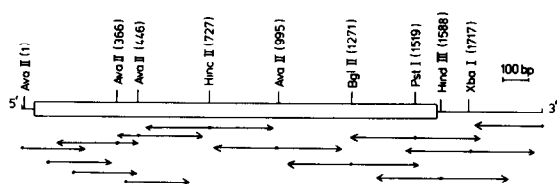


Fig.1. Partial restriction maps and sequencing strategy of the cDNA insert of λ cALI 23. Arrows mark the restriction sites sequenced from and the extent of the individual sequence determinations.

procedure and analyzed further by Southern blot hybridization and restriction mapping [12].

The inserts chosen for sequence analysis were subcloned in M13 mp18 and mp19 and sequenced according to Sanger and Coulson [13]. Full-length ALDH I cDNA was subcloned downstream of a

T₃-RNA-polymerase promotor [14] and transcribed in vitro with and without capping reagent (0.1 mM GpppG). The transcription reaction mixture was placed on ice and 10 μ l aliquots were directly translated without further purification in a rabbit reticulocyte lysate in the presence of [³⁵S]methionine. Proteins synthesized were separated on an SDS-polyacrylamide gel and detected by fluorography.

3. RESULTS AND DISCUSSION

40 hybridization signals were obtained after screening of 2×10^6 plaques of the human cDNA library with a nick-translated ALDH I cDNA fragment derived from pEXAL21 [3,9]. 24 signals were processed through three additional rounds of screening until distinct plaques could be isolated.



Fig.2. Nucleotide sequence of the cDNA insert of λ cALI 23 as determined by the strategy outlined in fig.1.

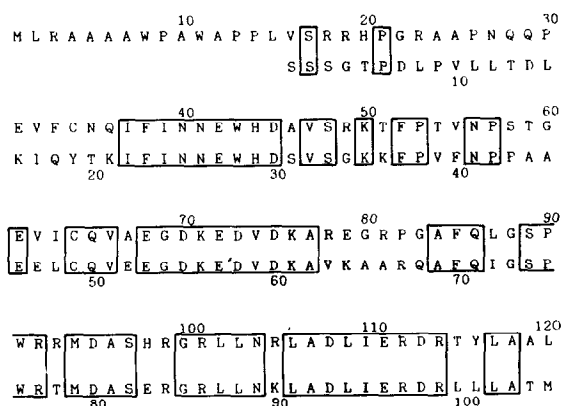


Fig.3. Comparison of the first 100 amino acids at the amino-terminal ends of ALDH I (upper sequence) and ALDH II (lower sequence).

Two clones with an approximate length of 2 kb were sequenced and found to be identical. Fig.1 shows a limited restriction map of the clone λ cALI 23 and the sequence strategy. The nucleotide sequence of the cDNA insert of λ cALI 23, which corresponds to the entire coding region of ALDH I mRNA and the adjacent non-coding regions, is shown in fig.2. The nucleotide sequence is in good agreement with a previously published partial sequence [15]. The deduced ALDH I amino acid sequence was subsequently compared to the known sequence of cytoplasmic isozyme ALDH II [2] in order to define the regions forming the mitochondrial signal peptide. As expected considerable homology (70%) was observed in the major part of the enzyme with exception of the amino-terminal end of both proteins (fig.3). The first 20 amino

acids of ALDH II showed no sequence homology to the corresponding ALDH I sequences. Furthermore, an additional peptide of 15 residues without a corresponding counterpart in ALDH II was found at the amino-terminal end of ALDH I. Based on these observations the whole of the 35 amino acid amino-terminal end of ALDH I seems to represent a mitochondrial leader peptide. Similar to other mitochondrial leader sequences (fig.4) this peptide is characterized by lack of acidic residues and an above average content of basic residues typically separated from each other by uncharged residues [6,7]. Whether all 35 amino acids belong to the leader sequence remains to be established. Recently, Hurt et al. [16] demonstrated that the first 12 amino acids of an imported mitochondrial protein are sufficient to direct a cytoplasmic protein into the mitochondrial matrix, although most leader peptides sequenced so far comprise 20–60 residues. In order to assess the actual length of ALDH I signal peptide, in vitro studies using recombinant ALDH I protein are essential. In pursuance of this goal, the whole cDNA insert of λ cALI 23 was subcloned into the vector pSP6/T3. This construct was transcribed in vitro by T₃-RNA-polymerase, and the resulting capped and uncapped mRNAs were translated in a cell-free reticulocyte lysate system. The synthesized polypeptide migrated similarly to authentic ALDH I on SDS-PAGE indicating the absence of major posttranslational modifications (fig.5). This protein product has an apparent molecular mass of 50 kDa which is in good agreement with the molecular mass of ALDH I monomers [1]. To define the critical regions in the putative leader peptide, in vitro targeting studies with modified ALDH I precursor are in progress.

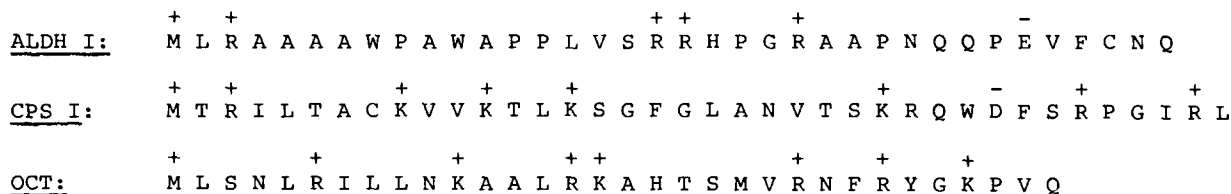


Fig.4. Comparison of ALDH I leader peptide and presequences found in other mitochondrial precursor proteins. (+) Basic amino acid, (–) acidic residue. OCT, ornithine carbamyltransferase, CPS I, carbamyl-phosphate synthetase I, ALDH I, aldehyde dehydrogenase I.

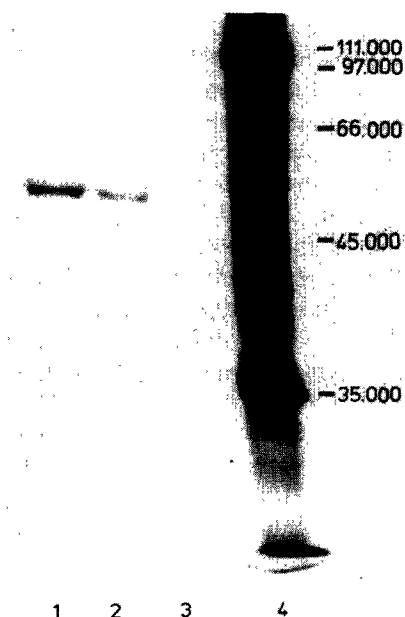


Fig.5. In vitro transcription-translation of the precursor of ALDH I. A fluorogram of the gel is shown. Molecular mass markers are indicated (in kDa). Lanes: 1, translation products of uncapped ALDH I mRNA; 2, translation products of capped ALDH I mRNA; 3, without exogenous RNA; 4, translation products of 5 μ g brome mosaic virus RNA.

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